Cytowish Ltd
3 Rue John Ernest Dolibois, 4573
Oberkorn Differdange, Luxembourg
Email: info@cytowish.com
Web: www.cytowish.com

Your authentic partner in molecular cytogenetics

BCOR(Xp11.4) Gene Break Apart Probe Detection Kit

(CW-128)

Intended use

This kit performs fluorescence in situ hybridization staining on the basis of conventional staining, and provides auxiliary information for diagnosis for physicians. The test results are for clinical reference only and should not be used as the sole basis for clinical diagnosis. Clinicians should make comprehensive judgments on the test results based on factors such as the patient's condition, drug indications, treatment response and other laboratory test indicators.

Product composition

The kit consists of BCOR dual-color probe (100μL/Tube).

Storage condition

Keep sealed away from light at -20°C±5°C, and the validity period is 12 months.

After the cover is opened, it can be sealed and stored in 2~8℃ away from light within 24 hours. After the cover is opened, it should be sealed and stored in -20±5℃ away from light for a long time. Transport with temperature below 0°C.

Applicable instruments

Fluorescence microscopy imaging systems, including fluorescence microscopy and filter sets suitable for DAPI (367/452), Green (495/517), and Orange (547/565).

Sample requirements

1. Applicable specimen types: Paraffin-embedded specimens from surgical excision or biopsy.
2. The tissue should be fixed with 4% neutral formaldehyde solution within 1 hour after isolation. After tissue fixation, it is routinely dehydrated and embedded in paraffin.

Pretreatment

Baking: Slides heating at 80°C for 30min or 65°C for 2h or overnight.

Dewaxing: According to the customer laboratory protocol (Commonly with Xylene for 15min).

Cytowish Ltd
3 Rue John Ernest Dolibois, 4573
Oberkorn Differdange, Luxembourg
Email: info@cytowish.com
Web: www.cytowish.com

Hydration: Take out the slides and put them respectively into 100%, 85% and 70% EtOH at room temperature for 3 minutes each.

Take out the slides, and immerse them in deionized water for 3 minutes. Remove the excess of water on the slides by air-drying. Permeation: Immerse the slides in deionized water at 100°C and boil continuously for 20-40 minutes (Conventional 20min). Remove the excess of water on the slides by air-drying.

Digestion: Protease enzymic digestion at 37°C for 10-40 minutes. Mix the protease work buffer (50mmol HCl) and the 10x protease solution (Pepsin concentration 5%) in a proportion of 9:1 to prepare the enzymatic digestion solution.

Washing: Wash with 2xSSC at room temperature for 5 minutes.

Dehydration: Take out the slides and dehydrate in 70%, 85%, and 100% gradient ethanol at room temperature for 2 minutes each time. Remove the excess of EtOH solution on the slides by air-drying.

Denaturing hybridization

The following operations should be carried out in the dark room.

Cell sample

1. Take out the probe, leave it at room temperature for 5min, turn it upside down with force, mix it well, and then centrifuge it for a short time (no vortex instrument vibration). Take 10μL of it and drop it into the cell drop hybridization area, immediately cover the cover glass of 22mm × 22mm. The probe should be evenly expanded under the cover glass without bubbles, and seal the edge with rubber glue (the edge must be completely sealed to prevent the dry piece from affecting the test results in the hybridization process).
2. The cell drops were placed on the hybridizer and denatured at 85°C for 2min (the hybridizer should be preheated to 85°C) and hybridized at 42°C for 2 to 16 hours.

Washing

The following operations should be performed in a darkroom.

1. Carefully tear off the adhesive around the cover glass with tweezers to avoid sticking off or moving the cover glass. Immerse the cell drop into 2xSSC for about 5s, and take it out. Gently push one corner of the cover glass to the edge of the slide with tweezers, and gently remove the cover glass with tweezers.

Cytowish Ltd
3 Rue John Ernest Dolibois, 4573
Oberkorn Differdange, Luxembourg
Email: info@cytowish.com
Web: www.cytowish.com

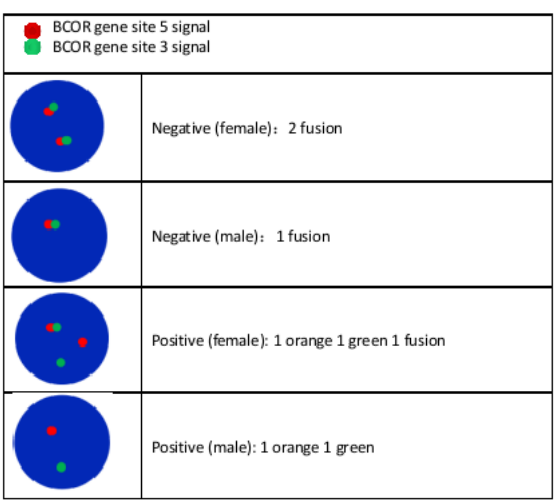
2. The cells were placed at 2xSSC room temperature for 1min.
3. 3% NP-40/0.4 xSSC solution preheated at 68℃for 2min.
4. The sides were immersed in deionized water preheated at 37℃ for 1min, and then dried naturally in the dark.
5. **Counterstaining**

The following operations should be performed in a darkroom.

10μL DAPI compound dye is dropped in the hybridization area of the glass slide and immediately covered. The suitable filter is selected for glass slide observation under the fluorescence microscope.

FISH results observation

P lace the stained sections under a fluorescence microscope and the cells area is first confirmed under a low magnification objective (10×); under magnification objective (40×) a uniform cells distribution is observed; then the nucleus size uniformity, nuclear boundary integrity, DAPI staining uniformity, no nuclei overlapping, cells clear signal are observed in the high magnification objective (100x).



Precautions

1. Please read this manual carefully before testing. The testing personnel shall receive professional technical training. The signal counting personnel must be able to observe and distinguish orange red and green signals.
2. When testing clinical samples, if it is difficult to count the hybridization signals and the samples are not enough to repeat the retest, the test will not provide any test

Cytowish Ltd
3 Rue John Ernest Dolibois, 4573
Oberkorn Differdange, Luxembourg
Email: info@cytowish.com
Web: www.cytowish.com

results. If the amount of cells is insufficient for analysis, again, the test will not provide test results.

3. The formamide and DAPI counterstaining agent used in this experiment have potential toxicity or carcinogenicity, so they need to be operated in the fume hood and wear masks and gloves to avoid direct contact.
4. The results of this kit will be affected by various factors of the sample itself, but also limited by enzyme digestion time, hybridization temperature and time, operating environment and limitations of current molecular biology technology, which may lead to wrong results. The user must understand the potential errors and accuracy limitations that may exist in the detection process.
5. All chemicals are potentially dangerous. Avoid direct contact. Used kits are clinical wastes and should be properly disposed of.
6. This product is for clinical diagnosis and scientific research.

[Manuscript version and approval date]

Manual version: V1.2 revised on 07 December 2022

Approval date: April 30, 2020